

## Review

---

# Algae: A potent antioxidant source

Neelma Munir<sup>1\*</sup>, Nadia Sharif<sup>1</sup>, Shagufta Naz<sup>1</sup> and Farkhanda Manzoor<sup>2</sup>

<sup>1</sup>Department of Biotechnology and Microbiology, Lahore College for Women University, Lahore.

<sup>2</sup>Department of Zoology, Lahore College for Women University, Lahore.

Accepted 23 April, 2013

---

**There is a current worldwide interest in finding new and safe antioxidants from natural sources. Algae can biosynthesize, metabolize, accumulate and secrete a great diversity of primary and secondary metabolites including carotenoids, phenolic compounds, phycobilins, sulphated compounds and vitamins. All of these compounds have commercial applications in medicine, pharmaceutical, nutraceutical, cosmetic, agricultural and food industries. Keeping in view the presence of these antioxidant compounds there is a great potential of algae that can be used in pharmaceuticals and other industries.**

**Key words:** Algae, antioxidants, carotenoids, photosynthesis, phycocyanin, vitamins.

---

## INTRODUCTION

Algae are photosynthetic organisms which constitute a total of 25 to 30 species, with a great diversity of forms and sizes. They can exist from unicellular microscopic organisms (microalgae) to multicellular of great size (macroalgae). Algae can be an interesting natural source of novel compounds with biological activity. Some algae are organisms that live in complex habitats submitted to extreme environmental conditions (salinity, temperature, nutrients, UV-Vis irradiation, etc.). To survive in these adverse conditions, they must adapt rapidly by producing a great variety of secondary (biologically active) metabolites, which cannot be found in other organisms (Gonulol et al., 2009; Welker et al., 2012). Among the various compounds found in the algae, antioxidants are probably the substances that have attracted major interest of the health and pharmaceutical industry (Table 1).

Substance that effectively prevents or delays the adverse effects caused by free radicals, even when the amount of the substance is less than the substance to be

oxidized is generally defined an antioxidant. Commercial antioxidant supplements such as butylated hydroxyl anisole (BHA), butylated hydroxyl toluene (BHT),  $\alpha$ -tocopherol and propyl gallate have been used to reduce oxidative damages (Gulçin et al., 2002). However, it is suspected that these synthetic antioxidants are responsible for a number of side-effects such as liver damage and carcinogenesis. As a result, consumers have become more health conscious and investigators are increasingly seeking natural antioxidant alternatives for use in foods or medicinal materials. Recently many researchers are interested in finding natural antioxidants having safety and effectiveness, which can be substituted the synthetic antioxidants, BHA and BHT. One such natural source for producing antioxidants is algae (Hirata et al., 2000; Benedettia et al., 2004; Karawita et al., 2007; Lee et al., 2008; Moo-Puc et al., 2011 ).

## Algal antioxidants

As algae are photosynthetic organisms, they produced free radicals and other oxidative reagents when they are exposed to high oxygen concentrations and light. It is considered because of absence of structural damage that these organisms are able to generate the necessary compounds to protect themselves against oxidation

---

\*Corresponding author. E- mail: [neelma.munir@yahoo.com](mailto:neelma.munir@yahoo.com).  
Tel.: +923004381090.

**Abbreviations:** BHA, butylated hydroxyl anisole; BHT, butylated hydroxyl toluene.

**Table1.** Antioxidant compounds from algae and their health benefits.

S. No	Antioxidant compounds	Perceived Health Benefits	Reference
1.	$\beta$ -carotene , lutein	Protective against breast cancer	(Yang et al., 2010)
2.	Bromophenol Carrageenan,  oligosaccharides Fucoidan	$\alpha$ glycosidase inhibition Anti - tumor  Anti – HIV  Ameliorates hyperoxaluria Anticancer Protection against neurogenerative disorder	(Kim et al., 2010) (Haijin et al., 2003) (Moo-Puc et al., 2011)  (Veena et al., 2007) (Luo et al., 2009)
3.	Fucophlorethols	Chemopreventive	(Parys et al., 2010)
4.	Fucoxanthin	Antiangiogenic Protective effects against retinol deficiency	(Sugawara et al., 2006) (Sangeetha et al., 2009)
5.	Galactan sulphate	Antiviral	(Talarico et al., 2004) (Yasuhara – Bell and Lu 2010)
6.	Phenolic functional groups and MAAs (mycosporine – like amino acids)	Antiproliferative	(Yuan et al., 2005)
7.	Phlorotannins	Anti – inflammatory Bactericide Inhibits H <sub>2</sub> O <sub>2</sub> mediated DNA damage Hypertension Photochemopreventive effect	(Shin et al., 2006) (Nagayama et al., 2002) (Ahn et al., 2007)  (Cha et al., 2006) (Hwang et al., 2006)
8.	Phycocerythrin	Amelioration of diabetic complications	(Yabuta et al., 2010)
9.	Polyphenols	Vascular chemoprotection Antimicrobial $\alpha$ glycosidase inhibition	(Kang et al., 2003) (Apostolidis and Lee 2010) (Jimenez et al., 2010)
10.	Porphyran, shinorine	Antiaging	(Zhang et al., 2003;Rastogi et al., 2010)

hence, algae is a potent antioxidant compounds that could also be suitable for protecting our bodies against the damaging effect of reactive oxygen species produced as a result of normal metabolism of the body. Algal antioxidants are of very different nature such as carotenoids and vitamin E ( $\alpha$ -tocopherol) are fat-soluble fraction, whereas vitamins, phycobiliproteins and polyphenols are the powerful water-soluble antioxidants (Table 2) (Ikeda et al., 2003; Ina Kamei 2006).

### Carotenoids

Carotenoids are among the most common pigments in nature. Most naturally occurring carotenoids are hydrophobic tetraterpenoids that contain a C40 methyl-branched hydrocarbon backbone (Figure 1). The polyene chains of carotenoids, consisting of conjugated double bonds, are responsible for the pigmentation of carotenoids and their ability to absorb photons in visible

wavelengths. Carotenoids are synthesized by all photosynthetic organisms as well as by many non-photosynthetic bacteria and fungi. More than 600 known carotenoids were reported in nature and about 50 have provitamin- A activity (Del Campo et al., 2007). There are two main classes of naturally occurring carotenoids: carotenes, which are hydrocarbons that are either linear or cyclized at one or both ends of the molecule (such as  $\beta$ - carotene,  $\alpha$ -carotene); and xanthophylls, which are oxygenated derivatives of carotenes. All xanthophylls produced by higher plants, for example violaxanthin, antheraxanthin, zeaxanthin, neoxanthin and lutein are also synthesized by green algae. However, in contrast to land plants, specific green algae possess additional xanthophylls such as loroxanthin astaxanthin and canthaxanthin (Baroli and Niyogi 2000; Grunewald et al., 2001). In addition, diatoxanthin, diadinoxanthin, and fucoxanthin are produced in brown algae and dinoflagellates (Lohr and Wilhelm, 2001).

In the algae the carotenoids seem to function primarily

**Table 2.** Antioxidant compounds in algae and potential algal sources for utilization.

Sr. no	General category	Example compounds	Algal source	Reference
1.	Carotenoids	$\beta$ -carotene	<i>Chondrus crispus</i> <i>Mastocarpus stellatus</i> Brown algae Red algae	Lohrmann et al. (2004)
2.	Phenolic compounds	Lycopene Stypodiol, Isoepitaondiol, Taondiol Terpenoids	Taonia atomaria	Nguyen and Schwartz (1999) Nahas et al. (2007)
3.	Phycobilin pigments	Phycoerythrin, phycocyanin	<i>Cystoseira</i> sp. Red algae in general	Foti et al. (1994) Romay et al. (2003); Sekar and Chandramohan (2008); Soni et al. (2009); Yabuta et al. (2010)
4.	Polyphenols	Catechin, Epicatechin, Gallate Flavonoids Phlorotannins	<i>Halimeda</i> sp.	Devi et al. (2008)
5.	Sulphated polysaccharides	Fucoidan, Alginic acid, Laminaran Fucoidan Sulphated galactans (lambda carrageenan) Galactans Sulphated Glycosaminoglycan Porphyran	<i>Turbinaria conoides</i>  <i>Laminaria japonica</i> Most red algae  Some marine red algae <i>Sargassum</i> <i>wightii</i> <i>Porphyra</i> sp.	Chattopadhyay et al. (2010)  Luo et al. (2009) Rocha de Souza et al.(2007) Barahona et al. (2011) Costa et al. (2010) Josephine et al. (2008) Athukorala et al. (2006)
6.	Vitamins	Ascorbate  Vitamin A	<i>Chondrus crispus</i> <i>Mastocarpus stellatus</i> <i>Sargassum</i> sp. <i>Kappaphycus</i> <i>alvarezii</i>	Lohrmann et al. (2004)  García-Casal et al. (2009) Kumar et al. (2008)

as photoprotective agents and as accessory light harvesting pigment, thereby protecting the photosynthetic apparatus against photo damage. They also play a role in phototropism and phototaxis. Some microalgae can undergo a carotenogenesis process, in response to various environmental and cultural stresses (e.g. light, temperature, salts, nutrients), where the alga stops growth and changes dramatically its carotenoid metabolism, accumulating secondary carotenoids as an adaptation to severe environments (Bhosale, 2004).

The main carotenoids produced by microalgae are  $\beta$ -carotene from *Dunaliella salina* and astaxanthin from *Haematococcus pluvialis*.  $\beta$ -carotene serves as an essential nutrient and has high demand in the market as a natural food colouring agent, as an additive to cosmetics and also as a health food (Raja et al., 2007).  $\beta$ -carotene is routinely used in soft-drinks, cheeses and

butter or margarines. Is well regarded as being safe and indeed positive health effects are also ascribed to this carotenoids due to a pro-vitamin A activity (Baker and Gunther, 2004).

Lycopene is a red pigment that occurs naturally in certain fruits, vegetables, algae and fungi. It belongs to a large group of pigments known as carotenoids; however, it has no provitamin A activity (Bradbury et al., 2012).

The benefits of astaxanthin are said to be numerous, and include enhancing eye health, improving muscle strength and endurance and protecting the skin from premature ageing, inflammation and UVA damage, is a strong coloring agent and has many functions in animals such as growth, vision, reproduction, immune function, and regeneration. Some reports support the assumption that daily ingestion of astaxanthin may protect body tissues from oxidative damage as this might be a



(*Arthrospira*) for phycocyanin (blue) and the rhodophyte *Porphyridium* for phycoerythrin (red). Phycocyanin is currently used in Japan and China as a natural colouring, in food products like chewing gums, candies, dairy products, jellies, ice creams, soft drinks (e.g. Pepsi® blue and also in cosmetics such as lipsticks, eyeliners and eye shadows. In a recent study, phycocyanin was considered a more versatile blue colorant than gardenia and indigo, providing a bright blue color in jelly gum and coated soft candy, despite its lower stability towards heat and light (Jespersen et al., 2005).

### Phenolic and polyphenols compounds

Polyphenolic secondary metabolites are a large and diverse group of chemical compounds which exist both in terrestrial plants and in aquatic macrophytes. Phenolic compounds can act as antioxidants by chelating metal ions, preventing radical formation and improving the antioxidant endogenous system (Al-Azzawie and Mohamed-Saiel, 2006). The term “phenolic compound” describes several hundred molecules found in edible plants that possess on their structure a benzenic ring substituted by, at least, one hydroxyl group (Manach et al., 2004). These phenolic compounds are commonly found in plants, including seaweeds (Duan et al., 2006). Polyphenols represent a diverse class of compounds including flavonoids (i.e. flavones, flavonols, flavanones, flavononols, chalcones and flavan-3-ols), lignins, tocopherols, tannins and phenolic acids (Thomas and Kim, 2011). Phlorotannins from brown algae have up to eight interconnected rings. They are therefore more potent free radical scavenger than other polyphenols derived from terrestrial plants, including green tea catechins, which only have three to four rings (Hemat, 2007).

Flavonoids, the largest groups of phenolic compounds are known to contain a broad spectrum of chemical and biological activities including antioxidant and free radical scavenging properties (Kahkonen et al., 1999). Flavonoids include flavonols, flavones, catechins, proanthocyanidins, anthocyanidins and isoflavonoids (Ndhlala et al., 2007).

In the brown algae (Phaeophyceae), the only group of tannins present is the phlorotannins. They are polymers of phloroglucinols (1,3,5-trihydroxybenzene, and may constitute up to 15% of the dry weight of brown algae (Wang et al., 2012).

### Sulphated polysaccharides

Polysaccharides are the most abundant among the

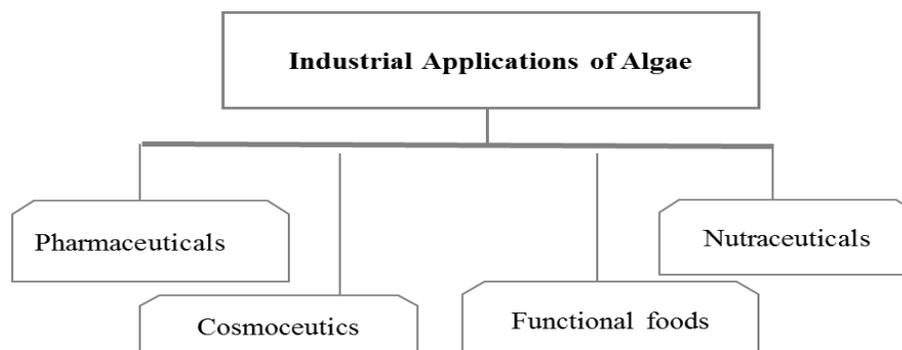
natural products produced by plants and they widely exist in plants, animals, microorganisms and algae ( Paulsen, 2002; Yang and Zhang 2009). These polymeric carbohydrate structures, usually composed of various monosaccharides linked with different glucosidic bonds (Holdt and Kraan 2011). Depending on the structure, polysaccharides can have distinct functional properties from their building blocks. Sulphated polysaccharides are among the most abundant and broadly studied polysaccharides from non-animal origin (Pereira et al., 2002). Most naturally occurring sulphated polysaccharides are complex mixtures of molecules showing wide variations in their structure as well as their activities (Alban et al., 2002). They are widespread in nature. Seaweeds are recognized as a major source of sulphated polysaccharides with various biological activities. Thus, sulphated polysaccharides are of special interest in the search of natural products.

Fucoidan is a type of complex sulphated polysaccharides, mainly found in the cell-wall matrix of various brown seaweed species (Teruya et al., 2007; O’Connell et al., 2008; Kim et al., 2010). It contains substantial percentages of L-fucose and sulfate ester groups (Matou et al., 2002; Li et al., 2008; Jiang et al., 2010). Fucose is a hexose deoxy sugar with the chemical formula  $C_6H_{12}O_5$  and is the fundamental sub unit of the fucoidan polysaccharides. For the past decade fucoidan has been extensively studied due to its numerous biological activities. Recently researches for new drugs have raised interest in fucoidans. In the past few years, several fucoidans’ structures have been isolated, and many aspects of their biological activity have been elucidated (Aisa et al., 2005; Li et al., 2008).

### Vitamins and minerals

Extraction of vitamins from natural sources has received increasing attention due to the high antioxidant activity associated to this family of compounds. Besides its well-known antioxidant activity, recent studies have demonstrated that certain synthetic vitamins are less effective than natural vitamins (Kelman et al., 2012).

Microalgae biomass represents a valuable source of nearly all essential vitamins (e.g. A, B1, B2, B6, B12, C, E, nicotinate, biotin folic acid and pantothenic acid) and a balanced mineral content (e.g. Na, K, Ca, Mg, Fe, Zn and trace minerals) (Becker, 2004). The high levels of vitamin B12 and Iron in some microalgae, like *Spirulina*, makes them particularly suitable as nutritional supplements for vegetarian individuals. The vitamin content of an alga depends on the genotype, the stage in the growth cycle, the nutritional status of the alga, the light intensity (photosynthetic rate). The vitamin content is



**Figure 2.** Industrial applications of algae.

therefore amenable to manipulation by varying the culture conditions as well as by strain selection or genetic engineering. However, vitamins cell content fluctuates with environmental factors, the harvesting treatment and the biomass drying methods (Garcia et al., 2009).

## POTENTIAL APPLICATIONS OF ANTIOXIDANTS

Algae are an enormous biological resource, representing one of the most promising sources for new products and industrial applications (Figure 2) (Pulz and Gross, 2004). They can be used to enhance the nutritional value of food and animal feed, due to their well-balanced chemical composition. Moreover, they are cultivated as a source of highly valuable molecules such as polyunsaturated fatty acids, pigments, antioxidants, pharmaceuticals and other biologically active compounds. The application of microalgal biomass and/or metabolites is an interesting and innovative approach for the development of healthier food products. Microalgal biotechnology is similar to conventional agriculture, but has received quite a lot of attention over the last decades, because they can reach substantially higher productivities than traditional crops and can be extended into areas and climates unsuitable for agricultural purposes (e.g. desert and seashore lands). Microalgae production is an important natural mechanism to reduce the excess of atmospheric CO<sub>2</sub> by biofixation and recycling of fixed C in products, ensuring a lower greenhouse effect, reducing the global environmental heating and climate changes. Microalgae cultivation also presents less or no seasonality, are important as feed to aquaculture and life-support systems, and can effectively remove nutrients (or pollutants) (e.g nitrogen and phosphorus) from water. Microalgal systems for sunlight driven environmental and production applications can clearly contribute to sustainable development and improved management of

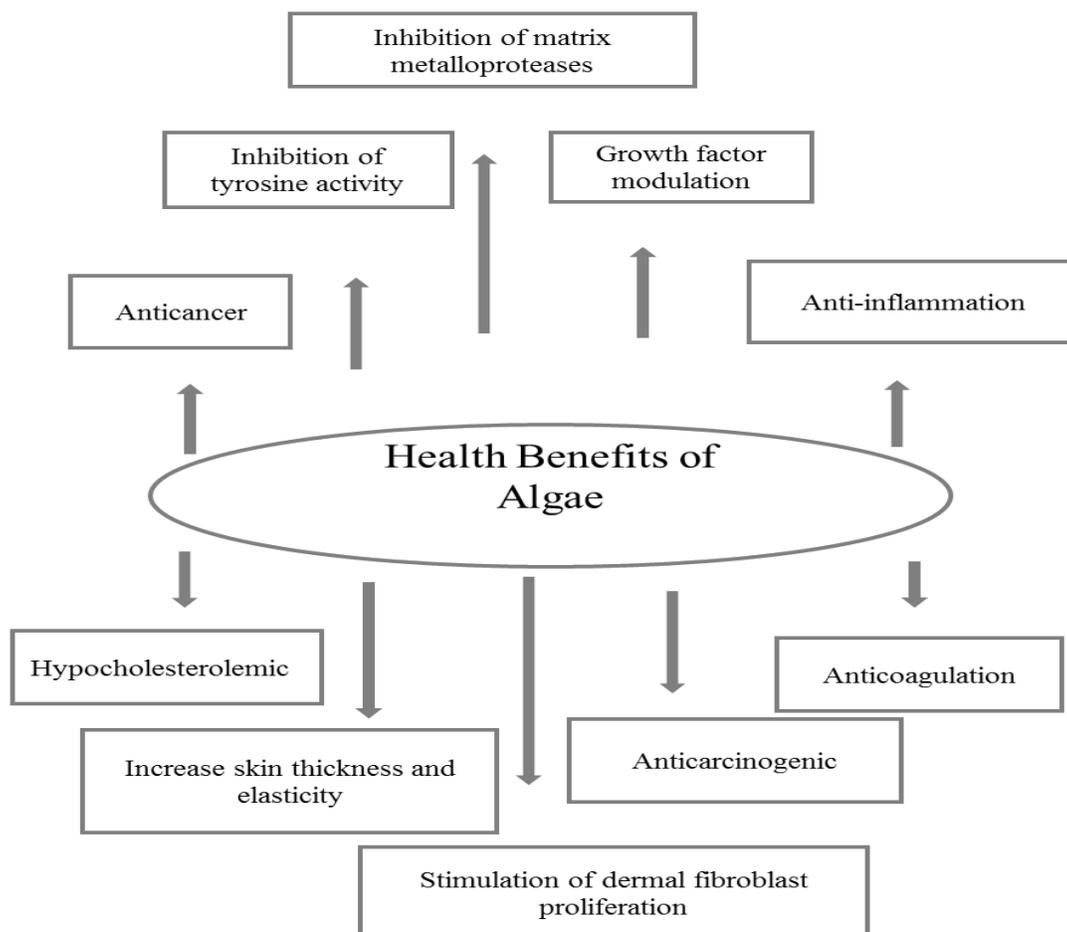
natural resources. Lately, microalgae have been seen with a great potential as a sustainable feedstock for biodiesel production, in substitution for oil from vegetable crops (Spolaore et al., 2006), and also for hydrogen production (Dutta et al., 2005).

## Human health

The United States Food and Drug Administration approved the use of astaxanthin as a feed additive for aquaculture in 1987 and subsequently in 1999 astaxanthin were approved as a nutraceutical (Guerin et al., 2003).

## Nutrition

In recent years, there has been a growing interest in the so-called functional foods because they can provide physiological benefits additional to nutritional and energetic, as, for instance, antihypertensive, antioxidant or anti-inflammatory. In the search for feasible new sources of natural antioxidants that can be used in the food industry, algae have been suggested as possible raw materials. These organisms are widely known and consumed in certain countries, and numerous health benefits have been associated with their use. Therefore, algae are potentially a great source of natural compounds that could be used as ingredients for preparing functional foods. Different compounds with antibacterial, antiviral and anti fungal activity can be found in these types of organisms along with compounds with antioxidant activity (Molina et al., 2003; Christaki et al., 2011; Lee et al., 2013). Antioxidants can play an important role in food technology because of their usefulness against lipid peroxidation (Senorans et al., 2003).



**Figure 3.** Health benefits of algae.

All over the world commercial production of microalgae for human nutrition is already a reality. Numerous combinations of microalgae or mixtures with other health foods can be found in the market in the form of tablets, powders, capsules, pastilles and liquids, as nutritional supplements (Table 2). They can also be incorporated into food products (e.g. pastas, biscuits, bread, snack foods, candies, yoghurts, soft drinks), have beneficial health effects that are associated with microalgal biomass, probably related to a general immunomodulating effect. In spite of some reluctance for novel foods in the past, nowadays there is an increasing consumer demand for more natural food products, presenting health benefits. Functional foods supplemented with microalgae biomass are sensorially much more convenient and variable, thus combining health benefits with attractiveness to consumers. The pathway of health benefits of algae is shown in Figure 3

(Pulz and Gross, 2004).

### Live stock

Using even very small amounts of micro algal biomass can positively affect the physiology of animals by improved immune response, resulting in growth promotion, disease resistance, antiviral and antibacterial action, improved gut function, probiotic colonization stimulation, as well as by improved feed conversion, reproductive performance and weight control. The external appearance of the animals may also be improved, resulting in healthy skin and a lustrous coat, for both farming animals (poultry, cows, breeding bulls) and pets (cats, dogs, rabbits, ornamental fishes and birds). Since feed corresponds to the most important exogenous factor influencing animal health and also the

major expense in animal production, the use of alternative high quality protein supplements replacing conventional protein sources is encouraged. Considering that animal feed stands at the beginning of the food chain, increasing public and legislative interest is evident, especially considering intensive breeding conditions and the recent trend to avoid “chemicals” like antibiotics (Breithaupt, 2007). The large number of nutritional and toxicological evaluations already conducted has demonstrated the suitability of algae biomass as a valuable feed supplement. In fact, 30% of the current world algal production is sold for animal feed applications (Becker, 2004).

## Cosmetics

Components of algae are frequently used in cosmetics as thickening agents, water-binding agents and antioxidants. Cosmetics companies claim benefits on the skin or health in general from contents like carrageenan, other algal polysaccharides, algal proteins or lipids, vitamin A, vitamin B1, iron, phosphorus, sodium, copper, magnesium, calcium, or other elements; some companies promise that algal extracts inhibit oxidative degeneration of collagen and hyaluronic acid and that they have anti-aging properties. From a scientific point of view, many of the promised effects have to be judged as not scientifically proven and unsubstantiated. Typical species that are used for cosmetics are *Chondrus crispus*, *Mastocarpus stellatus*, *Laminaria* spp., *Porphyra* spp., *Ulva lactuca*, *Ascophyllum nodosum*, *Alaria esculenta*, *Spirulina platensis*, *Nannochloropsis oculata*, *Chlorella vulgaris* and *Dunaliella salina* (Hallmann, 2007).

## Conclusion

Natural antioxidants, found in algae, are important bioactive compounds that play an important role against various diseases and aging processes through protection of cells from oxidative damage. It is a valuable source of neuroprotective agents. It provides a useful alternative to non - natural substances. Additionally the combination of the exceptional nutritional value of microalgae with colouring and therapeutically properties, associated with an increase demand of natural products, make microalgae worth exploring for utilization in the future in feed, food, cosmetic and pharmaceutical industries, with recognized advantages comparing with the traditional ingredients.

## REFERENCES

Aisa Y, Miyakawa Y, Nakazato T, Shibata H, Saito K, Ikeda Y,

- Kizaki M (2005). Fucoidan induces apoptosis of human HS-Sultan cells accompanied by activation of caspase-3 and down-regulation of ERK pathways. *Am. J. Hematol.*, 78: 7-14.
- Ahn GN, Kim KN, Cha SH, Song CB, Lee J, Heo MS, Yeo IK, Lee NH, Jee YH, Kim JS, Heu MS Jeon YJ (2007). Antioxidant activities of phlorotannins purified from *Ecklonia cava* on free radical scavenging using ESR and H<sub>2</sub>O<sub>2</sub> mediated DNA damage. *Eur. Food. Res. Technol.*, 226: 71-79.
- Alban S, Schauerte A, Franz G (2002). Anticoagulant sulfated polysaccharides. Part I. Synthesis and structure-activity relationships of new pullulan sulfates. *Carb. Polym.*, 47: 267-276.
- Al-Azzawie HF, Mohamed-Saiel SA (2006). Hypoglycemic and antioxidant effect of oleuropein in alloxan-diabetic rabbits. *Life Sci.*, 78: 1371-1377.
- Apostolidis E, Lee CM (2010). *In vitro* potential of *Ascophyllum nodosum* phenolic antioxidant-mediated alpha-glucosidase and alpha-amylase inhibition. *J. Food. Sci.*, 75: 97-102.
- Athukorala Y, Kim KN, Jeon YJ (2006). Antiproliferative and antioxidant properties of an enzymatic hydro-lysate from brown alga, *Ecklonia cava*. *Food. Chem. Toxicol.*, 44: 1065-1074.
- Becker EW (2004). Microalgae in human and animal nutrition. In: Richmond A, ed, *Handbook of microalgal culture*, Oxford, Blackwell, pp. 312-351.
- Baker R, Gunther C (2004). The role of carotenoids in consumer choice and the likely benefits from their inclusion into products for human consumption. *Trends. Food. Sci. Technol.*, 15: 484-488.
- Barahona T, Chandia NP, Encinas MV, Matsuhiro B, Zúñiga EA (2011). Antioxidant capacity of sulfated poly-saccharides from seaweed: a kinetic approach. *Food Hydrocoll.*, 25: 529 - 535.
- Baroli I, Niyogi KK (2000). Molecular genetics of xanthophylls-dependent photoprotection in green algae and plants. *Phil. Trans. R. Soc. Lond. B.* 355: 1385- 1394.
- Benedettia S, Benvenutia F, Pagliarania S, Francoglia S, Scogliob S, Canestraria F (2004). Antioxidant properties of a novel phycocyanin extract from the blue-green alga *Aphanizomenon flos-aquae*. *Life Sci.*, 75: 2353-2362.
- Bhosale P (2004). Environmental and cultural stimulants in the production of carotenoids from microorganisms. *Appl. Mic. Biotech.*, 63: 351-361
- Bradbury LM, Shumskaya M, Tzfadia O, Wu SB, Kennelly EJ, Wurtzel ET (2012) Lycopene cyclase paralog CruP protects against reactive oxygen species in oxygenic photosynthetic organisms. *Proc. Natl. Acad. Sci., U.S.A.* 109 (27):15.
- Breithaupt DE (2007). Modern application of xanthophylls in animal feeding - a review. *Tre. F. Sci. Technol.*, 18: 501-506.
- Chattopadhyay N, Ghosh T, Sinha S, Chattopadhyay K, Karmakar P, Ray B (2010). Polysaccharides from *Turbinaria conoides*: structural features and antioxidant capacity. *Food. Chem.*, 118:823-829.
- Cha SH, Lee KW, Jeon YJ (2006). Screening of extracts from red algae in Jeju for potential marine angiotensin-I converting enzyme (ACE) inhibitory activity. *Alg.*, 21: 343-348.
- Christaki E, Florou-Paneri P, Bonos E (2011). Microalgae: a novel ingredient in nutrition. *Int. J. food. Sci. Nutr.*, 62(8) :794-799.
- Costa LS, Fidelis GP, Cordeiro SL, Oliveira RM, Sabry DA, Câmara RBG, Nobre LTDB, Costa, MSSP, Almeida-Lima J, Farias EHC, Leite EL, Rocha HAO (2010). Biological activities of sulfated polysaccharides from tropical seaweeds. *Biomed. Pharm.*, 64: 21-28.
- Del Campo JA, Garcia-Gonzalez M, Guerrero MG (2007). Outdoor cultivation of microalgae for carotenoid production: current state and perspectives. *Appl. microbial biotechnol.*, 74: 1163-1174.
- Devi KP, Suganthy N, Kesika P, Pandian SK (2008). Bio-protective properties of seaweeds: in vitro evaluation of antioxidant activity and antimicrobial activity against food borne bacteria in relation to polyphenolic content. *BMC Complement Altern. Med.*, 8: 38.
- Dufosse L, Galaup P, Yaron A, Arad SM, Blanc P, Murthy KNC, Ravishankar, GA (2005). Microorganisms and microalgae as sources of pigments for food use: a scientific oddity or an industrial reality?. *Trends. Food Sci. Technol.*, 16: 389-406.
- Dutta D, De D, Chaudhuri S, Bhattacharya SK (2005). Hydrogen production by Cyanobacteria. *Microbial, Cell. Fact.* 4: 36-46.

- Duan XJ, Zhang WW, Li XM, Wang BG (2006). Evaluation of antioxidant property of extract and fractions obtained from a red alga, *Polysiphonia urceolata*. Food. Chem., 95: 37- 43.
- Garcia-Casal MN, Ramirez J, Leets I, Pereira AC, Quiroga MF (2009). Antioxidant capacity, polyphenol content and iron bioavailability from algae (*Ulva* sp., *Sargassum* sp. and *Porphyra* sp.) in human subjects. Br. J. Nutr., 101: 79-85.
- Gonulol A, Ersanli E, Baytul O (2009). Taxonomical and numerical comparison of epipelagic algae from Balik and Uzun lagoon, Turkey. J. Environ. Biol. 30: 777-784.
- Grunewald K, Hirschberg J, Hagen C (2001). Ketocarotenoid biosynthesis outside of plastids in the unicellular green alga *Haematococcus pluvialis*. J. Biol. Chem., 276: 6023- 6029.
- Gulçin I, Oktay M, Kufrevioglu OI, Aslan A (2002). Determination of antioxidant activity of lichen *Cetraria islandica* (L). Ach. J. Ethnopharmacol., 79: 325-329.
- Guerin M, Huntley ME, Oliazola M (2003). *Haematococcus* astaxanthin: Applications for human health and nutrition. Trends. Biotechnol., 21: 210-216.
- Haijin M, Xiaolu J, Huashi G (2003). A k-carrageenan derived oligosaccharide prepared by enzymatic degradation containing anti-tumor activity. J. Appl. Phycol., 15: 297-303.
- Hallmann A (2007). Algal transgenics and biotechnology. Transg. Plant J., 1: 81- 98.
- Hemat RAS (2007). Fat and muscle dysfunction. In R. A. S. Hemat eds, Andropathy. Dublin, Ireland: Urotext, pp. 83-85.
- Hirata T, Tanaka M, Ooike M, Tsunomura T, Sakaguchi M (2000). Antioxidant activities of phycocyanobilin prepared from *Spirulina platensis*. J. Appl. Phycol., 12: 435-439.
- Holdt SL, Kraan S (2011). Bioactive compounds in seaweeds: functional food applications and legislation. J. Appl. Phycol., 23: 543-597.
- Hwang H, Chen T, Nines RG, Shin HC, Stoner GD (2006). Photochemoprevention of UVB-induced skin carcinogenesis in SKH-1 mice by brown algae polyphenols. Int. J. Canc., 119: 2742-2749.
- Ina A, Kamei Y (2006). Vitamin B<sub>12</sub>, a chlorophyll-related analog to pheophytin a from marine brown algae, promotes neurite outgrowth and stimulates differentiation in PC12 cells. Cytotechnol., 52: 181-187.
- Ikeda K, Kitamura A, Machida H, Watanabe M, Negishi H, Hiraoka J, Nakano T. (2003). Effect of *Undaria pinnatifida* (Wakame) on the development of cerebrovascular diseases in stroke prone spontaneously hypertensive rats. Clin. Experim. Pharmacol. Physiol., 30: 44-48.
- Jespersen L, Stromdahl LD, Olsen K, Skibsted, LH (2005). Heat and light stability of three natural blue colorants for use in confectionery and beverages. Eur. Food. R. Technol., 220: 261-266.
- Jiang Z, Okimura T, Yokose T, Yamasaki Y, Yamaguchi K, Oda T (2010). Effect of sulfated, ascophyllan, from the brown alga *Ascophyllum nodosum* on various cell lines: A comparative study on ascophyllan and fucoidan. J. Biosci. Bioeng., 110: 113-117.
- Jimenez JT, O'Connell S, Lyons H, Bradley B, Hall M (2010). Antioxidant, antimicrobial, and tyrosinase inhibition activities of acetone extract of *Ascophyllum nodosum*. Chem. Pap., 64: 434-442.
- Josephine A, Nithya K, Amudha G, Veena CK, Preetha SP, Varalakshmi P (2008). Role of sulphated polysaccharides from *Sargassum Whightii* in Cyclosporine A-induced oxidative liver injury in rats. BMC Pharmacol., 8: 4.
- Karawita R, Senevirathne M, Athukorala Y, Affan A, Lee YJ, Kim SK, Lee JB, Jeon YJ (2007). Protective effect of enzymatic extracts from microalgae against DNA damage induced by H<sub>2</sub>O<sub>2</sub>. Marine Biotechnol., 9: 479-490.
- Kahkonen MP, Hopia AI, Vuorela HJ, Rauha JP, Pihlaja K, Kujala TS (1999). Antioxidant activity of plant extracts containing phenolic compounds. J. Agric. Food. Chem., 47: 3954-3962.
- Kang K, Park Y, Hwang HJ, Kim SH, Lee JG, Shin HC (2003). Antioxidative properties of brown algae polyphenolics and their perspectives as chemopreventative agents against vascular risk factors. Arch. Pharm. Res., 26: 286-293.
- Kelman D, Posner EK., McDermid K.J, Tabandera NK, Wright PR, Wright AD (2012) Antioxidant activity of Hawaiian marine algae. Marine Drugs. 10: 403-416.
- Kim K, Lee OH, Lee BY (2010). Fucoidan, a sulfated polysaccharide, inhibits adipogenesis through the mitogen-activated protein kinase pathway in 3T3-L1 preadipocytes. Life Sci., 86: 791-797.
- Kumar KS, Ganesan K, Subba Rao PV (2008). Antioxidant potential of solvent extracts of *Kappaphycus alvarezii* (Doty) Doty: an edible seaweed. Food. Chem., 107: 289-295.
- Lee SH, Karawita R, Affan A, Lee JB, Lee BJ, Jeon YJ (2008). Potential antioxidant activities of enzymatic digests from benthic diatoms *Achnanthes longipes*, *Amphora coffeaeformis*, and *Navicula* sp. (*Bacillariophyceae*). J. Food. Sci. Nutr., 13: 166-175.
- Lee K.Y, Jeong MR, Choi SM, Na SS, Cha, JD (2013) Synergistic effect of fucoidan with antibiotics against oral pathogenic bacteria. Archiv. Oral. Biol., 58: 482-492.
- Li B, Lu F, Wei X, Zhao R (2008). Fucoidan: Structure and bioactivity. Molecule. 13: 1671-1695.
- Lohr M, Wilhelm C (2001). Xanthophyll synthesis in diatoms: Quantifications of putative intermediate and comparison of pigment conversion kinetics with rate constants derive from a model. Planta. 212: 382- 391.
- Lohrmann NL, Logan BA, Johnson AS (2004). Seasonal acclimatization of antioxidants and photosynthesis in *Chondrus crispus* and *Mastocarpus stellatus*, two co-occurring red algae with differing stress tolerances. Biol. Bull., 207: 225-232.
- Luo D, Zhang Q, Wang H, Cui Y, Sun Z, Yang J, Zheng Y, Jia J, Yu F, Wang X, Wang X (2009). Fucoidan pro-TECTS against dopaminergic neuron death *in vivo* and *in vitro*. Eur. J. Pharmacol., 617: 33-40.
- Manach C, Scalbert A, Morand C, Remes C, Jimenez L (2004). Polyphenols: food sources and bioavailability. A. J. Clin. Nutr., 79: 727-747.
- Matou S, Helley D, Chabut D, Bros A, Fischer AM (2002). Effect of fucoidan on fibroblast growth factor-2-induced angiogenesis *in vitro*. Thrombosis Res., 106: 213-221.
- Molina Grima E, Belarbi EH, Acien Fernandez FG, Robles Medina A, Chisti Y (2003). Biotechnol. Adv., 20: 491.
- Moo-Puc R, Robledo D, Freile-Pelegrin Y (2011). Enhanced Antitumoral Activity of Extracts Derived from Cultured *Udotea flabellum* (Chlorophyta). Evid. Based. Complement. Alternat. Med., 969275: 21.
- Nagayama K, Iwamura Y, Shibata T, Hirayama I, Nakamura T (2002). Bactericidal activity of phlorotannins from the brown alga *Ecklonia kurome*. J. Antimicrob. Chemother., 50: 889-893.
- Nahas R, Abatis D, Anagnostopoulou MA, Kefalas P, Vagias C, Roussis V (2007). Radical-scavenging activity of Aegean Sea marine algae. Food. Chem., 102: 577-581.
- Ndhkala AR, Kasiyamhuru A, Mupure C, Chitindingu K, Benhura MA, Muchuweti M (2007). Phenolic composition of *Flacourtia indica*, *Opuntia megacantha* and *Sclerocarya birrea*. Food Chem., 103: 82-87.
- O'Connell E, Murray P, Piggot C, Hennequart F, Tuohy, M (2008). Purification and characterization of a N-acetylglucosaminidase produced by *Talaromyces emersonii* during growth on algal fucoidan. J. Appl. Phycol., 20: 557-565.
- Pereira MS, Silva ACESV, Valente AP, Mourao P AS (2002). A 2-sulfated, 3-linked -l-galactan is an anticoagulant polysaccharide. Carb. Res., 337: 2231-2238.
- Parys S, Kehraus S, Krick A, Glombitza KW, Carmeli S, Klimo K, Gerhäuser C, König GM (2010). *In vitro* chemopreventive potential of fucophlorethols from the brown alga *Fucus vesiculosus* by antioxidant activity and inhibition of selected cytochrome P450 enzymes. Phytochem., 71: 221-229.
- Paulsen BS (2002). Biologically active polysaccharides as possible lead compounds. Phytochem. Rev., 1: 379-387.
- Pulz O, Gross W (2004). Valuable products from biotechnology of microalgae. Appl. Microbiol. Biotechnol., 65: 635-648.
- Raja R, Hemaiswarya S, Rengasamy R (2007). Exploitation of *Dunaliella* for  $\alpha$ -carotene production. Appl. Microbiol. Biotechnol., 74: 517-523.

- Rastogi RP, Richa Sinha RP, Singh SP, Hader DP (2010). Photoprotective compounds from marine organisms. *J. Ind. Microbiol. Biotechnol.*, 37: 537-558.
- Rocha de Souza MC, Marques CT, Dore CMG, Fer-reira da Silva FR, Rocha HAO, Leite E L. (2007). Antioxidant activities of sulfated polysaccharides from brown and red seaweeds. *J. Appl. Phycol.*, 19: 153-160.
- Romay C, Gonzalez R, Ledon N, Ramirez D, Rimbau V (2003). C-phycoerythrin: a biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Curr. Protein Pept. Sci.*, 4: 207-216.
- Sangeetha RK, Bhaskar N, Baskaran V (2009). Comparative effects of  $\beta$ -carotene and fucoxanthin on retinol deficiency induced oxidative stress in rats. *Mol. Cell. Biochem.*, 331: 59-67.
- Sekar S, Chandramohan, M (2008). Phycobiliproteins as a commodity: trends in applied research, patents and commercialization. *J. Appl. Phycol.*, 20: 113-136.
- Senorans FJ, Ibanez E, Cifuentes (2003) A Critical Review. *Food. Sci. Nutr.*, 43: 507.
- Shin HC, Hwang HJ, Kang KJ, Lee BH (2006). An antioxidative and anti-inflammatory agent for potential treatment of osteoarthritis from *Ecklonia cava*. *Arch. Pharm. Res.*, 29: 165-171.
- Soni B, Visavadiya NP, Madamwar D (2009). Attenuation of diabetic complications by C-phycoerythrin in rats: antioxidant activity of C-phycoerythrin including copper-induced lipoprotein and serum oxidation. *Br. J. Nutr.*, 102: 102-109.
- Spolaore P, Joannis Cassan C, Duran E, Isambert A (2006). Commercial applications of Microalgae- review. *J. Biosci. Bioeng.*, 101: 87-96.
- Sugawara T, Matsubara K, Akagi R, Mori M, Hirata T (2006). Antiangiogenic activity of brown algae fucoxanthin and its deacetylated product, fucoxanthinol. *J. Agri. Food. Chem.*, 54: 9805-9810.
- Talarico LB, Zibetti RGM, Faria PCS, Scolaro LA, Duarte MER, Noseda MD, Pujol CA, Damonte EB (2004). Anti-herpes simplex virus activity of sulfated galactans from the red seaweeds *Gymnogongrus griffithsiae* and *Cryptonemia crenulata*. *Int. J. Biol. Macromol.*, 34: 63-71.
- Teruya T, Konishi T, Uechi S, Tamaki H, Tako M (2007). Anti-proliferative activity of oversulfated fucoidan from commercially cultured *Cladosiphon okamuranus* TOKIDA in U937 cells. *Int. J. Biol. Macromol.*, 41: 221-226.
- Thomas NV, Kim SK (2011). Potential pharmacological applications of polyphenolic derivatives from marine brown algae. *Environ. toxicol. pharmacol.*, 32: 325-335.
- Van den Berg H, Faulks R, Granado HF, Hirschberg J, Olmedilla B, Sandmann G, Southon S, Stahl W. (2000). The potential for the improvement of carotenoid levels in foods and the likely systemic effects. *J. Sci. Food. Agric.*, 80: 880-912.
- Veena CK, Josephine A, Preetha SP, Varalakshmi P (2007). Beneficial role of sulfated polysaccharides from edible seaweed *Fucus vesiculosus* in experimental hyperoxaluria. *Food. Chem.*, 100: 1552-1559.
- Wang T, Jonsdottir R, Liu H, Gu L, Kristinsson HG, Raghavan S, Olafsdottir G (2012). Antioxidant Capacities of Phlorotannins Extracted from the Brown Algae *Fucus vesiculosus*. *J. Agric. Food. Chem.*, 60:5874-5883.
- Welker M, Dittmann E, von Dohren H (2012). Cyanobacteria as a source of natural products. *Methods. Enzymol.* 517: 23-46.
- Yabuta Y, Fujimura H, Kwak CS, Enomoto T, Wata-nabe, F. (2010). Antioxidant activity of the phycoerythrin-rubilin compound formed from a dried Korean purple laver (*Porphyra* sp.) during *in vitro* digestion. *Food. Sci. Technol. Res.*, 16: 347-351.
- Yasuhara Bell J, Lu Y. (2010). Marine compounds and their antiviral activities. *Antivir Res.*, 86: 231-240.
- Yang L, Zhang LM. (2009). Chemical structure and chain conformational characterization of some bioactive polysaccharides isolated from natural sources. *Carb. Polym.*, 76: 349-361.
- Yang YJ, Nam SJ, Kong G, Kim M K. (2010). A case control study on seaweed consumption and the risk of breast cancer. *Br. J. Nutr.*, 103: 1345-1353.
- Yuan YV, Carrington MF, Walsh, NA. (2005). Extracts from dulse (*Palmaria palmata*) are effective antioxidants and inhibitors of cell proliferation *in vitro*. *Food. Chem. Toxicol.*, 43: 1073-1081.
- Zhang Q, Li N, Zhou G, Lu X, Xu Z, Li Z. (2003). *In vivo* antioxidant activity of polysaccharide fraction from *Porphyra haitanensis* (Rhodophyta) in aging mice. *Pharmacol. Res.*, 48: 151-155.